

The essential guide

Multiomic single cell immunology

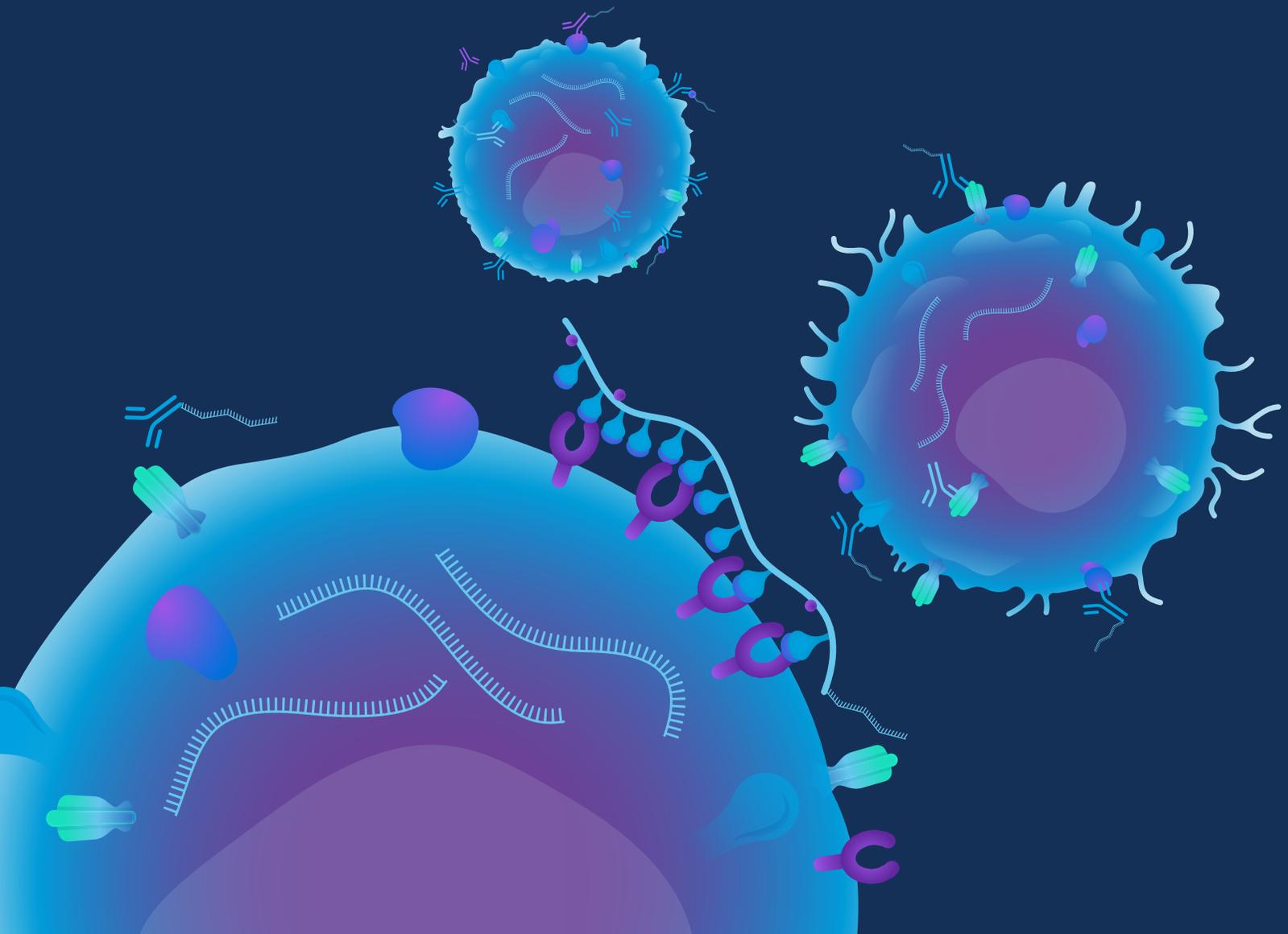


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Section 1

Introduction

Immunologists have long relied on methods that provide analysis of individual immune cells to understand the immune system's diversity and complexity. While technologies such as flow or mass cytometry have been pivotal for answering complex questions in immunology, continuing advances in the field will require the integration of multiomic single cell data.

Single cell multiomics for immunology provides access to multiple readouts at once for thousands to millions of single cells, giving you more from your single cell analyses. Imagine, instead of getting one measurement—protein expression—you could get everything at once: protein and gene expression, T- and B-cell receptor clonotypes, and antigen specificity; or coupled transcriptomic and epigenetic data. Instead of whittling down your markers of interest to a small panel that needs to be redesigned whenever new proteins are included, multiomic immunology lets you stain for all possible markers of interest at the same time, without the challenges of spectral overlap or compensation matrices. You can also combine immunophenotypic profiling and clonotype analysis with a readout of cell type and state for every cell. Single cell multiomic immunology means you can learn more from a single sample, without having to split it into parts, and improve reproducibility by profiling thousands of cells at a time and easily aggregating or comparing samples.

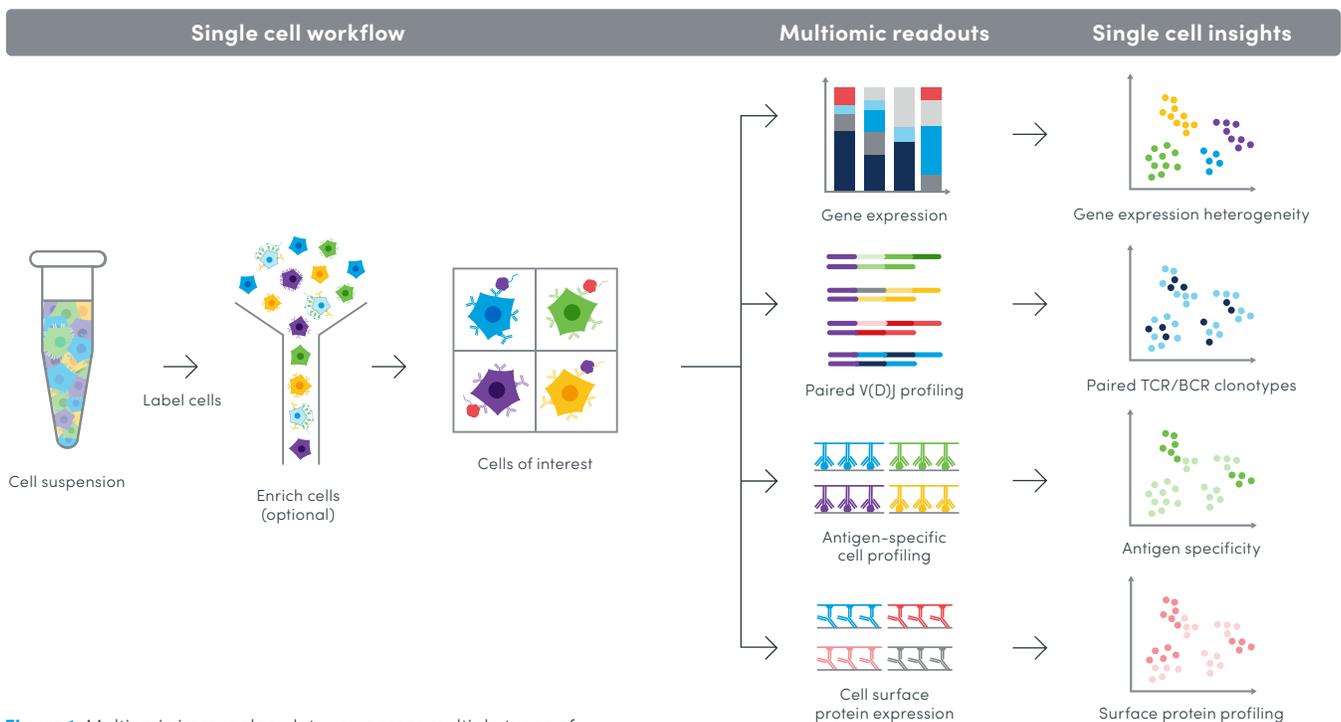


Figure 1. Multiomic immunology lets you access multiple types of information at once for thousands to millions of single cells.

Section 2

Explore what you can do with multiomic single cell immunology

Researchers have already started to answer a diverse set of immunological questions using single cell approaches. Here are some examples of immunology publications showing the unique ways these questions can be approached with single cell and spatial technology.

From what progenitor are my cells derived?

Combine single cell gene expression analysis with lineage tracing

Publications: [10](#), [36](#), [40](#)

What cell types are responsible for disease?

Discover rare cell types, subtypes, and novel functions

Publications: [24](#), [25](#), [27](#), [28](#), [81](#)

Understand cellular interactions that contribute to disease and identify novel therapeutic targets

Publications: [21](#), [60](#), [68](#), [75](#)

What genes are important for immune activation?

Screen hundreds of genetic edits simultaneously with single cell CRISPR screens

Publications: [14](#), [46](#), [47](#), [62](#)

How rampant is an ongoing infection?

Capture host and pathogen RNA transcripts from the same cells

Publications: [59](#), [60](#), [61](#), [66](#), [67](#)

How do I understand the immune response to vaccination or infection in patients?

Measure vaccine response

Publications: [7](#), [64](#), [73](#), [74](#), [87](#), [89](#)

Discover antibodies

Publications: [1](#), [4](#), [6](#), [8](#)

What dictates patient disease severity and response to therapy?

Identify expanded T-cell or B-cell receptor clonotypes

Publications: [19](#), [22](#), [67](#)

Identify immune factors that contribute to different disease response or prognosis

Publications: [71](#), [76](#), [77](#)

Track patient responses to cell therapy

Publication: [53](#)

Characterize immune cell activation state alongside clonotype and antigen binding specificity

Publications: [55](#), [63](#)

Identify the gene regulatory networks that govern T-cell exhaustion

Publication: [45](#)

Tools for high resolution multiomic immunology

Characterize complex cell populations by profiling hundreds of cell surface proteins along with gene expression and more, cell by cell, to gain an intricate picture of immunology and accelerate your discoveries with comprehensive solutions from 10x Genomics.

Targeted Gene Expression

Enrich sequencing libraries for your transcripts of interest using pre-designed or custom panels to focus on the genes that matter most.

Single Cell Immune Profiling

Get full-length, paired T- and B-cell receptor sequences, so you can match clonotype expansion to cell type and state, and generate functional receptors or antibodies for in vitro testing.

Multiomic Cytometry

Turn a next-generation sequencer into an ultra-high parameter cytometric detector to profile hundreds of protein markers at once, with optional transcriptome, T- and B-cell receptor sequences, or antigen specificity on top.

Single Cell ATAC

Survey the physical structure of the genome by identifying regions of open chromatin to reveal areas of active gene transcription, regulatory regions, and binding site motifs.

Single Cell Multiome ATAC + Gene Expression

Profile both the transcriptome and epigenome simultaneously in the same single cells, enabling deeper characterization of gene regulatory networks and cell state.

Single Cell CRISPR Screening

Directly assess CRISPR-driven gene edits or knockdowns and resulting gene expression phenotypes cell by cell to dissect molecular underpinnings of human immunity.

Single Cell Gene Expression

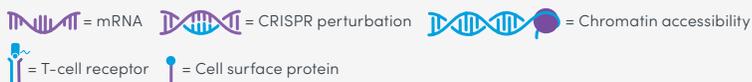
Obtain a digital readout of gene expression levels that lends insight into cellular heterogeneity, diversity, development, function, and response to external stimuli.

Immune Receptor Mapping

Characterize antigen specificity and phenotype with protein markers and/or gene expression to track vaccine response and accelerate antibody discovery.

Spatial Transcriptomics and Proteomics

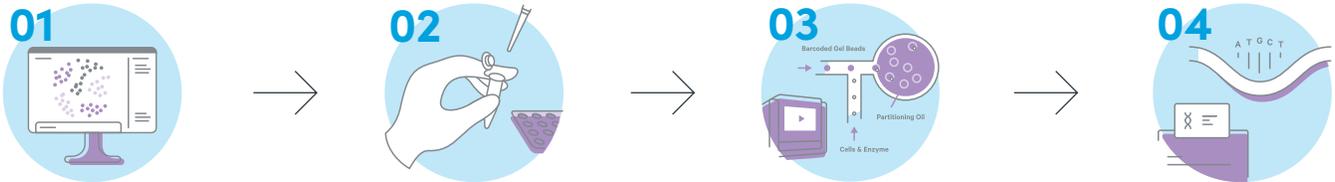
Visium [Spatial Gene Expression](#) and [Spatial Proteomics](#) provide transcriptome and protein readouts from intact tissue sections to complement or extend your single cell analyses.



Section 3

Planning your experiment

Before starting your single cell experiments, we recommend that you consider several key factors to help guide your experimental design and determine how to best answer your research questions.



01

Decide on the best single cell application

Depending on your biological question, different single cell approaches may be desired. If you want to study vaccine response or identify novel antibodies, [Single Cell Immune Receptor Mapping](#) can provide functional clonotype information and antigen binding specificity. For comprehensive phenotyping by gene expression and protein, [Multiomic Cytometry](#) may be your best choice. When you need to understand gene regulatory networks, [Single Cell ATAC](#) or [Single Cell Multiome ATAC + Gene Expression](#) can generate the data you require. Refer to the [Tools for high resolution multiomic immunology](#) section for a full list of single cell applications from 10x Genomics, or explore the product pages at [10xgenomics.com](https://www.10xgenomics.com).

02

Preparing samples

Sample type & preparation

It is critical that you obtain a clean single cell suspension free of cell debris, with minimal cell aggregates and high viability (>70%). It is also important to know the size range of the cells studied. The cell size is usually correlated with the number of transcripts expressed in the cell. A wide range of cell sizes (up to 30 μm) are compatible with Chromium Single Cell Next GEM Chips. In general, cell preparation protocols will vary depending on the tissue's origin and the cell types studied. Each tissue type is unique, and thus, it is critical to optimize sample preparation before starting any single cell experiment. We recommend starting by reviewing our [Cell Preparation Guide](#). Find more answers to common questions about sample prep on our [Sample Prep FAQs page](#). We also have sample prep-focused webinars available for viewing in our [Videos library](#).

Sample processing

The ability to process samples quickly after isolation or tissue dissociation is critical in maintaining cell integrity and preserving each cell's transcriptome. Be aware that any sample manipulations may adversely affect gene expression profiles, cell states, or cell viability and introduce bias into the study ([Van Den Brink et al., 2017](#)).

Cell enrichment

When characterizing rare cell populations, for applications such as immune receptor mapping or antibody discovery, enriching for cells of interest prior to generating single cell partitions can help ensure adequate numbers of your cells of interest. The Chromium system is compatible with [FACS](#) and [bead- or column-based](#) enrichment methods.

Cells versus nuclei

While some applications, such as Multiomic Cytometry or Immune Receptor Mapping, require a single cell suspension as input, others, including Single Cell ATAC and Single Cell Multiome ATAC + Gene Expression, require nuclei isolation. Cells are recommended for Single Cell Immune Profiling, but either cells or nuclei can be used for gene expression analysis with Single Cell Gene Expression.

Available sample types can also dictate whether cells or nuclei must be used. For frozen tissue or archival samples, nuclei must be [isolated](#) directly. Alternatively, tissue can be dissociated and [cryopreserved](#) immediately after it is received, enabling cells to be stored long term.

Species compatibility

Single cell gene expression products from 10x Genomics have been used successfully on a wide range of organisms. The capability to extend single cell gene expression analysis to other species may depend on the quality of the available reference annotation. Similarly, measuring cell surface proteins in other species requires effective antibody reagents with oligonucleotide conjugations. Targeted Gene Expression pre-designed or custom panels are provided only for human genes, although a small number of exogenous sequences, such as reporter genes, can be added. Single Cell Immune Profiling provides ready-made primers for amplification of human or mouse T- or B-cell receptor transcripts, but experimental design is flexible and other species have successfully been clonotyped using custom primers ([L Goldstein et al., 2019](#)).

03

Ensuring reliability

Number of cells

Deciding on the number of cells required depends on the expected cellular heterogeneity in the sample, the number of cells available, the minimum frequency expected of desired subpopulations, and the minimum number of cells of each cell type desired for data analysis (see online tool: satijalab.org/howmanycells). If the sample diversity is not known, a high number of cells at low sequencing depth may be the most flexible option to obtain a representative proportion of the cell population and meaningful biological information. Often, greater cell number, rather than sequencing depth, improves cell classification ability ([J Ding et al., 2020](#)). The Chromium system can recover up to ~65% of the cells loaded with a low doublet rate (0.9% per 1000 cells). For highly heterogeneous samples, thousands of cells may be required to fully resolve each subpopulation. However, the high cell recovery rate also makes Chromium suitable for samples with limited cell numbers.

Number of replicates

Determining the number of replicates depends on the research project, the type of sample, and the number of cells required in the study. The matter of biological replicates is still an [open question](#) in the field. In some studies, one sample alone can be seen as sufficient—with each cell representing a biological replicate, and different samples from different individuals accounting for the variability of a particular biological process. In other studies, to mitigate biological variability occurring in small cell populations across time, it can be beneficial to computationally aggregate cells from different samples to cover all aspects of the cell population being studied. Other cases may require the use of multiple replicates derived from a single sample to increase the total number of cells in the study.

Batch effects

Batch effects can be introduced at any stage of the workflow and are primarily due to logistical constraints that result in different preparation times, operators, and handling protocols. The 10x Genomics Chromium system demonstrates minimal [technical variability](#) across a variety of technical replicates. When combining data from multiple libraries, we recommend equalizing the sequencing read depth ([depth normalization](#)) between libraries before computationally merging to reduce batch effects introduced by sequencing. In addition, a number of computational tools including [Seurat](#), [scran](#), and [scrone](#) can correct batch effects. Cell Ranger can also perform batch correction for gene expression libraries.

04 Sequencing considerations

Sequencing depth

The sequencing depth per experiment for gene expression libraries is dependent on total mRNA content in individual cells, and the diversity of mRNA species. In general, at the same transcript diversity, cells expressing a low amount of mRNA will require less sequencing depth than cells expressing a large amount of mRNA. When sequencing cost or capacity is limiting, there is often a [trade-off](#) between sequencing a higher number of cells versus sequencing a lower number of cells with more reads, or [breadth versus depth](#). Single cell libraries for T- or B-cell receptor sequence, antigen specificity, protein markers, and CRISPR guides require less sequencing depth (minimum 5,000 read pairs per cell) than single cell ATAC (minimum 25,000 read pairs per cell) or single cell gene expression libraries (minimum 20,000 read pairs per cell).

10x Genomics single cell libraries are compatible with short-read sequencers and available in a dual indexing configuration. Our single cell gene expression workflows use unique molecular identifiers (UMIs) to barcode each transcript molecule before amplification takes place, resulting in a digital gene expression profile while accounting for PCR amplification bias.

Getting started with data analysis

Multimic immunology solutions from 10x Genomics come with intuitive software for data analysis and visualization.

Simultaneously measure single cell modalities

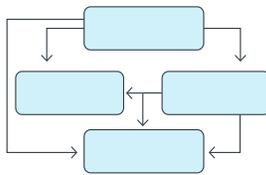
Gene expression
Cell surface protein

Full-length, paired V(D)J sequence
Antigen specificity

Chromatin accessibility
CRISPR perturbation



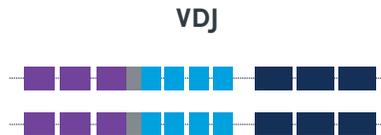
Go from DNA sequence to immunophenotype with a single software package



Cell Ranger is our suite of analysis pipelines that turn your raw sequencing data into results.

		Cell Barcodes				
Genes	0	3	0	0	...	
	0	0	0	0	...	
	0	0	0	0	...	
	2	0	0	5	...	
	:	:	:	:	...	

Cell Ranger provides a count matrix consisting of columns for every cell barcode and rows for all measured features, including genes for transcriptome analysis, protein markers, and bound antigens or pMHC multimers.



When applicable, Cell Ranger will also perform assembly and annotation of supported immune repertoire clonotypes, including full-length, paired V, D, and J sequences.



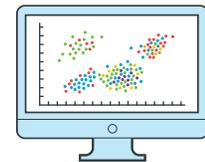
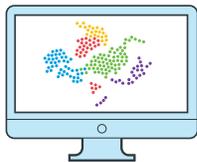
Interactively explore your multimic data

Loupe Browser lets you visualize and explore your results with point-and-click accessibility to:

Enable manual annotation of cell clusters

Import and overlay clonotype information

Re-cluster cells based on alternative features such as protein markers



Take your analysis further with third-party tools

Gene and protein expression data

Seurat, Scanpy, Bioconductor

Immune repertoire data

Immunarch, VDJ tools, scRepertoire

Working with a bioinformatician:

For some questions, it may be helpful to work with a bioinformatician specializing in single cell sequencing data. Sharing some biological context can help them zero in on data most relevant to your research question. For example, how many different cell types are expected? Were cells sorted before sequencing, so that one cell type should dominate the population? How many cells were targeted, and at what sequencing depth? Of course, specific data analysis questions can also be directed to the expert 10x Genomics Software Support team at support@10xgenomics.com.

Publications

From immune cell discovery to therapeutic development, there are already a wealth of publications highlighting the power of single cell immunology that can help guide your own investigations.

Antibody discovery

1. WB Alsoussi et al., A Potently Neutralizing Antibody Protects Mice against SARS-CoV-2 Infection. *J Immunol.* 205, 915–922 (2020).
2. T Bradley et al., RAB11FIP5 Expression and Altered Natural Killer Cell Function Are Associated with Induction of HIV Broadly Neutralizing Antibody Responses. *Cell.* 175, 387–399.e17 (2018).
3. Y Cao et al., Potent Neutralizing Antibodies against SARS-CoV-2 Identified by High-Throughput Single-Cell Sequencing of Convalescent Patients' B Cells. *Cell.* 182, 73–84.e16 (2020).
4. P Gilchuk et al., Integrated pipeline for the accelerated discovery of antiviral antibody therapeutics. *Nat Biomed Eng.* 4, 1030–1043 (2020).
5. L Goldstein et al., Massively parallel single-cell B-cell receptor sequencing enables rapid discovery of diverse antigen-reactive antibodies. *Commun Biol.* 2, 304 (2019).
6. F Horns, SR Quake, Cloning antibodies from single cells in pooled sequence libraries by selective PCR. *PLoS One.* 15, e0236477 (2020).
7. F Horns, CL Dekker, SR Quake, Memory B Cell Activation, Broad Anti-influenza Antibodies, and Bystander Activation Revealed by Single-Cell Transcriptomics. *Cell Rep.* 30, 905–913.e6 (2020).
8. I Setliff et al., High-Throughput Mapping of B Cell Receptor Sequences to Antigen Specificity. *Cell.* 179, 1636–1646.e15 (2019).

Assay method

9. A Dixit et al., Perturb-Seq: Dissecting Molecular Circuits with Scalable Single-Cell RNA Profiling of Pooled Genetic Screens. *Cell.* 167, 1853–1866.e17 (2016).
10. LS Ludwig et al., Lineage Tracing in Humans Enabled by Mitochondrial Mutations and Single-Cell Genomics. *Cell.* 176, 1325–1339.e22 (2019).
11. EP Mimitou et al., Multiplexed detection of proteins, transcriptomes, clonotypes and CRISPR perturbations in single cells. *Nat Methods.* 16, 409–412 (2019).
12. SA Overall et al., High throughput pMHC-I tetramer library production using chaperone-mediated peptide exchange. *Nat Commun.* 11, 1909 (2020).
13. VM Peterson et al., Multiplexed quantification of proteins and transcripts in single cells. *Nat Biotechnol.* 35, 936–939 (2017).
14. JM Replogle et al., Combinatorial single-cell CRISPR screens by direct guide RNA capture and targeted sequencing. *Nat Biotechnol.* 38, 954–961 (2020).
15. M Stoeckius et al., Simultaneous epitope and transcriptome measurement in single cells. *Nat Methods.* 14, 865–868 (2017).
16. SC Van Den Brink et al., Single-cell sequencing reveals dissociation-induced gene expression in tissue subpopulations. *Nat Methods.* 14, 935–936 (2017).
17. GXY Zheng et al., Massively parallel digital transcriptional profiling of single cells. *Nat Commun.* 8, 14049 (2017).

Autoimmunity

18. S Alivernini et al., Distinct synovial tissue macrophage subsets regulate inflammation and remission in rheumatoid arthritis. *Nat Med.* 26, 1295–1306 (2020).
19. BS Boland et al., Heterogeneity and clonal relationships of adaptive immune cells in ulcerative colitis revealed by single-cell analyses. *Sci Immunol.* 5, eabb4432 (2020).
20. N Borchering et al., A transcriptomic map of murine and human alopecia areata. *JCI Insight.* 5, e137424 (2020).
21. B Huang et al., Mucosal Profiling of Pediatric-Onset Colitis and IBD Reveals Common Pathogenics and Therapeutic Pathways. *Cell.* 179, 1160–1176.e24 (2019).
22. A Ramesh et al., A pathogenic and clonally expanded B cell transcriptome in active multiple sclerosis. *Proc Natl Acad Sci USA.* 117, 22932–22943 (2020).
23. D Schafflick et al., Integrated single cell analysis of blood and cerebrospinal fluid leukocytes in multiple sclerosis. *Nat Commun.* 11, 247 (2020).

Cell atlas

24. AE Gillen et al., Single-Cell RNA Sequencing of Childhood Ependymoma Reveals Neoplastic Cell Subpopulations That Impact Molecular Classification and Etiology. *Cell Rep.* 32, 108023 (2020).
25. W Guo et al., Single-cell transcriptomics identifies a distinct luminal progenitor cell type in distal prostate invagination tips. *Nat Genet.* 52, 908–918 (2020).
26. VM Howick et al., The Malaria Cell Atlas: Single parasite transcriptomes across the complete Plasmodium life cycle. *Science.* 365, eaaw2619 (2019).
27. D Montoro et al., A revised airway epithelial hierarchy includes CFTR-expressing ionocytes. *Nature.* 560, 319–324 (2018).
28. J Park et al., Single-cell transcriptomics of the mouse kidney reveals potential cellular targets of kidney disease. *Science.* 360, 758–763 (2018).
29. J-E Park et al., A cell atlas of human thymic development defines T cell repertoire formation. *Science.* 367, eaay3224 (2020).
30. JM Sà, et al., Single-cell transcription analysis of Plasmodium vivax blood-stage parasites identifies stage- and species-specific profiles of expression. *PLoS Biol.* 18, e3000711 (2020).
31. BJ Stewart et al., Spatiotemporal immune zonation of the human kidney. *Science.* 365, 1461–1466 (2019).
32. A Tikhonova et al., The bone marrow microenvironment at single-cell resolution. *Nature.* 569, 222–228 (2019).

Immunobiology

33. S Culemann et al., Locally renewing resident synovial macrophages provide a protective barrier for the joint. *Nature.* 572, 670–675 (2019).
34. JS Dahlin et al., A single-cell hematopoietic landscape resolves 8 lineage trajectories and defects in Kit mutant mice. *Blood.* 131, e1–e11 (2018).
35. J Ding et al., Characterisation of CD4+ T-cell subtypes using single cell RNA sequencing and the impact of cell number and sequencing depth. *Sci Rep.* 2, 304 (2020).
36. M Ghaedi et al., Single-cell analysis of RORα tracer mouse lung reveals ILC progenitors and effector ILC2 subsets. *J Exp Med.* 217, jem.20182293 (2020).
37. C Goudot et al., Aryl Hydrocarbon Receptor Controls Monocyte Differentiation into Dendritic Cells versus Macrophages. *Immunity.* 47, 582–596.e6 (2017).

38. JM Granja et al., Single-cell multiomic analysis identifies regulatory programs in mixed-phenotype acute leukemia. *Nat Biotechnol.* 37, 1458–1465 (2019).
39. T Hagai et al., Gene expression variability across cells and species shapes innate immunity. *Nature.* 563, 197–202 (2018).
40. L Kok et al., A committed tissue-resident memory T cell precursor within the circulating CD8+ effector T cell pool. *J Exp Med.* 217, e20191711 (2020).
41. T Luo et al., A single-cell map for the transcriptomic signatures of peripheral blood mononuclear cells in end-stage renal disease. *Nephrol Dial Transplant.* (2019). doi:10.1093/ndt/gfz227
42. G Mollaoglu et al., The Lineage-Defining Transcription Factors SOX2 and NKX2-1 Determine Lung Cancer Cell Fate and Shape the Tumor Immune Microenvironment. *Immunity.* 49, 764–779.e9 (2018).
43. L Pace et al., The epigenetic control of stemness in CD8+ T cell fate commitment. *Science.* 359, 177–186 (2018).
44. VS Patil et al., Precursors of human CD4+ cytotoxic T lymphocytes identified by single-cell transcriptome analysis. *Science Immunology.* 3, eaan8664 (2018).
45. AT Satpathy et al., Massively parallel single-cell chromatin landscapes of human immune cell development and intratumoral T cell exhaustion. *Nat Biotechnol.* 37, 925–936 (2019).
46. K Schumann et al., Functional CRISPR dissection of gene networks controlling human regulatory T cell identity. *Nat Immunol.* 21, 1456–1466 (2020).
47. E Shifrut et al., Genome-wide CRISPR Screens in Primary Human T Cells Reveal Key Regulators of Immune Function. *Cell.* 175, 1958–1971.e15 (2018).
48. M Shnayder et al., Defining the Transcriptional Landscape during Cytomegalovirus Latency with Single-Cell RNA Sequencing. *mBio.* 9, e00013–18 (2018).
49. PC Wilson et al., The single-cell transcriptomic landscape of early human diabetic nephropathy. *Proc Natl Acad Sci USA.* 116, 19619–19625 (2019).
50. D Zemmour et al., Single-cell gene expression reveals a landscape of regulatory T cell phenotypes shaped by the TCR. *Nat Immunol.* 19, 291–301 (2018).
51. Y Zeng et al., Single-Cell RNA Sequencing Resolves Spatiotemporal Development of Pre-thymic Lymphoid Progenitors and Thymus Organogenesis in Human Embryos. *Immunity.* 51, 930–948.e6 (2019).

Immunotherapies

52. MM Gubin et al., High-Dimensional Analysis Delineates Myeloid and Lymphoid Compartment Remodeling during Successful Immune-Checkpoint Cancer Therapy. *Cell.* 175, 1014–1030.e19 (2018).
53. Y-C Lu et al., Single-Cell Transcriptome Analysis Reveals Gene Signatures Associated with T-cell Persistence Following Adoptive Cell Therapy. *Cancer Immunol Res.* 7, 1824–1836 (2019).
54. K Paulson et al., Acquired cancer resistance to combination immunotherapy from transcriptional loss of class I HLA. *Nat Commun.* 9, 3868 (2018).
55. MR Rollins, EJ Spartz, IM Stromnes, T Cell Receptor Engineered Lymphocytes for Cancer Therapy. *Curr Protoc Immunol.* 129, e97 (2020).
56. A Sheih et al., Clonal kinetics and single-cell transcriptional profiling of CAR-T cells in patients undergoing CD19 CAR-T immunotherapy. *Nat Commun.* 11, 219 (2020).

Infectious disease

57. EA Stadtmauer et al., CRISPR-engineered T cells in patients with refractory cancer. *Science*. 367, eaba7365 (2020).
58. K Yost et al., Clonal replacement of tumor-specific T cells following PD-1 blockade. *Nat Med*. 25, (2019). P Bost et al. Host-Viral Infection Maps Reveal Signatures of Severe COVID-19 Patients. *Cell*. 181, 1475–1488.e12 (2020).
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61. V Cortez et al., Astrovirus infects actively secreting goblet cells and alters the gut mucus barrier. *Nat Commun*. 11, 2097 (2020).
62. Z Daniloski, Identification of required host factors for SARS-CoV-2 infection in human cells. *Cell*. (2020). doi:10.1016/j.cell.2020.10.030
63. AP Ferretti et al., Unbiased Screens Show CD8+ T Cells of COVID-19 Patients Recognize Shared Epitopes in SARS-CoV-2 that Largely Reside outside the Spike Protein. *Immunity*. 53, 1095–1107.e3 (2020).
64. Q Han et al., Neonatal Rhesus Macaques Have Distinct Immune Cell Transcriptional Profiles following HIV Envelope Immunization. *Cell Rep*. 30, 1553–1569.e6 (2020).
65. JS Lee et al., Immunophenotyping of COVID-19 and influenza highlights the role of type I interferons in development of severe COVID-19. *Sci Immunol*. 5, eabd1554 (2020).
66. R León-Rivera et al., Interactions of Monocytes, HIV, and ART Identified by an Innovative scRNAseq Pipeline: Pathways to Reservoirs and HIV-Associated Comorbidities. *mBio*. 11, e01037-20 (2020).
67. M Liao et al., Single-cell landscape of bronchoalveolar immune cells in patients with COVID-19. *Nat Med*. 26, 842–844 (2020).
68. E Park et al., *Toxoplasma gondii* infection drives conversion of NK cells into ILC1-like cells. *eLife*. 8, e47605 (2019).
69. M Reyes et al., An immune-cell signature of bacterial sepsis. *Nat Med*. 26, 333–340 (2020).
70. AB Russell, C Trapnell, JD Bloom, Extreme heterogeneity of influenza virus infection in single cells. *eLife*. 7, e32303 (2018).
71. Y Su et al., Multi-Omics Resolves a Sharp Disease-State Shift between mild and moderate COVID-19. *Cell*. (2020). doi: 10.1016/j.cell.2020.10.037
72. W Sungnak et al., SARS-CoV-2 entry factors are highly expressed in nasal epithelial cells together with innate immune genes. *Nat Med*. 26, 681–687 (2020).
73. A Waickman et al., Dissecting the heterogeneity of DENV vaccine-elicited cellular immunity using single-cell RNA sequencing and metabolic profiling. *Nat Commun*. 10, 3666 (2019).
74. AT Waickman et al., Transcriptional and clonal characterization of B cell plasmablast diversity following primary and secondary natural DENV infection. *EBioMedicine* 54, 102733 (2020).
75. W Wen et al., Immune cell profiling of COVID-19 patients in the recovery stage by single-cell sequencing. *Cell Discov*. 6, 31 (2020).
76. J-Y Zhang et al., Single-cell landscape of immunological responses in patients with COVID-19. *Nat Immunol*. 21, 1107–1118 (2020).

Inflammation and allergies

77. Y Zheng et al., A human circulating immune cell landscape in aging and COVID-19. *Protein & Cell*. 11, 740–770 (2020).
78. K Carlberg et al., Exploring inflammatory signatures in arthritic joint biopsies with Spatial Transcriptomics. *Sci Rep*. 9, 18975 (2019).
79. D Fernandez et al., Single-cell immune landscape of human atherosclerotic plaques. *Nat Med*. 25, 1576–1588 (2019).
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81. K Parikh et al., Colonic epithelial cell diversity in health and inflammatory bowel disease. *Nature*. 567, 49–55 (2019).
82. W Rao et al., Regenerative Metaplastic Clones in COPD Lung Drive Inflammation and Fibrosis. *Cell*. 181, 848–864.e18 (2020).
83. G Seumois et al., Single-cell transcriptomic analysis of allergen-specific T cells in allergy and asthma. *Sci Immunol*. 5, eaba6087 (2020).

Transplant

84. AJ Byrne et al., Dynamics of human monocytes and airway macrophages during healthy aging and after transplant. *J Exp Med*. 217, e20191236 (2020).
85. J Cai et al., Impact of Local Alloimmunity and Recipient Cells in Transplant Arteriosclerosis. *Circ Res*. 127, 974–993 (2020).
86. D Kim et al., Somatic mTOR mutation in clonally expanded T lymphocytes associated with chronic graft versus host disease. *Nat Commun*. 11, 2246 (2020).

Vaccine development

87. PS Arunachalam et al., T cell-inducing vaccine durably prevents mucosal SHIV infection even with lower neutralizing antibody titers. *Nat Med*. 26, 932–940 (2020).
88. E Kaufmann et al., BCG Educates Hematopoietic Stem Cells to Generate Protective Innate Immunity against Tuberculosis. *Cell*. 172, 176–190.e19 (2018).
89. Y Kotliarov et al., Broad immune activation underlies shared set point signatures for vaccine responsiveness in healthy individuals and disease activity in patients with lupus. *Nat Med*. 26, 618–629 (2020).

Resources from 10x Genomics

Application note

Discover how 10x Genomics technology is “Redefining Cellular Phenotyping” with this in-depth study of clonal expansion and antigen binding after viral infection.

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